## **Forum Editorial**

# Redox Regulation of Insulin Action and Signaling

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NSULIN is the major hormone contributing to glucose homeostasis. Aberrant secretion or action of this hormone has been implicated in the pathogenesis of diabetes mellitus (26). Insulin action on its target cells is initiated by insulininduced activation of intrinsic protein tyrosine kinase (PTK) activity of the  $\beta$  subunit by the insulin receptor (IR) (27). Once activated, IR-PTK can phosphorylate several cytosolic IR substrates (IRSs and Shc), which serve as docking sites for Src homology 2 domain-containing signaling molecules (25). Among these signaling molecules, phosphatidylinositol 3kinase (PI3-K) activation has emerged as a key player mediating the metabolic effects of insulin (22). PI3-K phosphorylates phosphatidylinositol (PI) lipids at position 3 of the inositol ring, and generates 3-phosphorylated forms of PI, such as phosphatidylinositol 3,4,5-trisphosphate, which are implicated in the activation of phospholipid-dependent kinases and related serine/threonine protein kinases responsible for the phosphorylation and stimulation of several downstream signaling protein kinases, such as protein kinase B (PKB) (also known as Akt), glycogen synthase kinase 3, p70s6k, and protein kinase C-ζ (22). Activation of these protein serine/threonine kinases has been demonstrated to play a key role in insulin-induced glucose transport, glucose transporter-4 (GLUT-4) translocation, glycogen, and protein

In recent years, considerable evidence has accumulated to suggest the involvement of reactive oxygen and nitrogen species (ROS/RNS) in modulating insulin signaling and action. Exogenous hydrogen peroxide ( $H_2O_2$ ) has been reported to mimic several physiological responses of insulin, such as glucose transport (9), glycogen synthesis (11), lipogenesis (14), lipolysis (12), and phosphoenolpyruvate carboxykinase gene expression (23), although the  $H_2O_2$  concentrations required for these effects were in the millimolar range (9, 11, 12, 14, 23). Furthermore, stimulation of cells with insulin was found to generate small amounts of  $H_2O_2$  (13) via a NADPH oxidase-dependent system (14). This insulin-induced, endogenously produced  $H_2O_3$  appears to be essential for initial tyro-

sine phosphorylation of IR and IRS protein, and is achieved by oxidative inhibition of protein tyrosine phosphatase (13, 14, 16). On the other hand, several studies have indicated that long-term exposure of various cell types to H<sub>2</sub>O<sub>2</sub> exerts an inhibitory effect on insulin action. For example, in L6 myotubes as well as in 3T3-L-1 adipocytes, H<sub>2</sub>O<sub>2</sub> treatment decreased insulin-induced glucose transport and membrane translocation of GLUT-4 (20). This effect was associated with attenuation of PI3-K/PKB signaling events and an increase in serine phosphorylation and degradation of IRS-1 (19). Phosphorylation of IRS-1 in specific serine residues has been implicated in dysregulated PI3-K/PKB activation and appears to be among the potential molecular mechanisms of insulin resistance (24, 26).

In vascular smooth muscle cells (VSMC) also, H<sub>2</sub>O<sub>2</sub> induced a decrease of PKB phosphorylation that was associated with a decline of IR autophosphorylation (6). In these studies, the total amount of IRS-1 remained unchanged by H<sub>2</sub>O<sub>2</sub> treatment (6), which is in contrast with reports on 3T3-L1 adipocytes or FAO rat hepatoma cells (19). However, these investigators did not examine if H<sub>2</sub>O<sub>2</sub> caused an increase in serine phosphorylation or a decrease in tyrosine phosphorylation of IRS-1 (6). Interestingly, pretreatment of VSMC with H<sub>2</sub>O<sub>2</sub> resulted in partial inhibition of insulin-binding activity in these cells (6). However, other studies, which have demonstrated an attenuation of insulin-induced responses by H<sub>2</sub>O<sub>2</sub> or other agents, have not reported a similar decrease of insulin-binding activity (8, 19, 20, 24). It thus appears that low levels of endogenously generated H2O2 function as an initiator (13, 14) or "primer" of IR phosphorylation (21) and subsequent downstream signaling, whereas higher concentrations and longer exposure to exogenous H2O2 serve as an attenuator of the insulin response (6, 8, 19, 20).

Another free radical, nitric oxide (NO), has also emerged as a modulator of insulin action. Enhanced NO production due to heightened inducible NO synthase (iNOS) activity has been suggested to contribute to insulin resistance in genetic and diet-induced rodent models of insulin resistance (18). NO

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has the ability to react to superoxide anion and to generate highly reactive peroxynitrite radicals that, by catalyzing the nitration of proteins, can impair their functions (1). In this regard, the peroxynitrite-induced nitration of several key tyrosine residues in IRS-1 was recently shown to reduce insulin stimulation of glucose uptake and the PI3-K/PKB signaling pathway (17). Further support for a potential role of iNOS as a negative modifier of insulin signaling and action is provided by studies in which iNOS-deficient mice were protected from diet-induced insulin resistance (18).

These developments implicating redox molecules as regulators of insulin signaling and action prompted this Forum issue of Antioxidants & Redox Signaling. The aim of this Forum is to highlight some important studies relating to the role of ROS and RNS in various aspects of insulin signaling and action. The issue contains one original contribution and six reviews in the field. The original article by Mehdi et al. explores the upstream components involved in H<sub>2</sub>O<sub>2</sub>-induced signaling events (15). The six reviews are written by experts in this important area of research, and summarize the work done in their own and other laboratories. Dröge discusses the mechanism and potential contribution of oxidative enhancement of IR autophosphorylation (3). Goldstein et al. describe the mechanism and targets of insulin-induced H<sub>2</sub>O<sub>2</sub> generation in initiating the insulin signaling cascade (7). The work presented by Konrad highlights the role of the antioxidant molecule α-lipoic acid as a stimulator/potentiator of the insulin signaling cascade and as a potential antidiabetic agent (10). Evans et al. summarize the mechanisms by which ROS/RNS contribute to insulin resistance (4). Frank et al. provide an up-to-date summary on the modulation of insulin signaling by redox in vascular and endothelial cells (5). Finally, Christon et al. discuss the recent progress made on the contribution of NO synthase in promoting insulin resistance and endothelial dysfunction (2).

I hope that this *Forum* issue will serve as a good source of information on redox modulation of insulin signaling and action. I wish to thank the authors for their excellent contributions, full cooperation, and sustained interest in putting this *Forum* issue together. I also appreciate the support from the Editors-in-Chief of *Antioxidants & Redox Signaling* for organizing this focused *Forum* issue.

### **ABBREVIATIONS**

GLUT-4, glucose transporter-4;  $\mathrm{H_2O_2}$ , hydrogen peroxide; iNOS, inducible nitric oxide synthase; IR, insulin receptor; IRS, insulin receptor substrate; NO, nitric oxide; PI, phosphatidylinositol; PI3-K, phosphatidylinositol 3-kinase; PKB, protein kinase B; PTK, protein tyrosine kinase; RNS, reactive nitrogen species; ROS, reactive oxygen species; VSMC, vascular smooth muscle cells.

#### REFERENCES

1. Beckman JS, Beckman TW, Chen J, Marshall PA, and Freeman BA. Apparent hydroxyl radical production by

- peroxynitrite: implications for endothelial injury from nitric oxide and superoxide. *Proc Natl Acad Sci U S A* 87: 1620–1624, 1990.
- Christon R, Drouin O, and Marette A. Redox modulation of insulin signaling and endothelial function. *Antioxid Redox Signal* 7: 1062–1070, 2005.
- Dröge W. Oxidative enhancement of insulin receptor signaling: experimental findings and clinical implications. *Antioxid Redox Signal* 7: 1071–1077, 2005.
- Evans JL, Maddux BA, and Goldfine ID. The molecular basis for oxidative stress-induced insulin resistance. Antioxid Redox Signal 7: 1040–1052, 2005.
- Frank GD, Eguchi S, and Motley ED. The role of reactive oxygen species in insulin signaling in the vasculature. Antioxid Redox Signal 7: 1053–1061, 2005.
- Gardner CD, Eguchi S, Reynolds CM, Eguchi K, Frank GD, and Motley ED. Hydrogen peroxide inhibits insulin signaling in vascular smooth muscle cells. *Exp Biol Med* (Maywood) 228: 836–842, 2003.
- Goldstein BJ, Mahadev K, Wu X, Zhu L, and Motoshima H. Role of insulin-induced reactive oxygen species in the insulin signaling pathway. *Antioxid Redox Signal* 7: 1021– 1031, 2005.
- Hansen LL, Ikeda Y, Olsen GS, Busch AK, and Mosthaf L. Insulin signaling is inhibited by micromolar concentrations of H<sub>2</sub>O<sub>2</sub>. Evidence for a role of H<sub>2</sub>O<sub>2</sub> in tumor necrosis factor alpha-mediated insulin resistance. *J Biol Chem* 274: 25078–25084, 1999.
- Hayes GR and Lockwood DH. Role of insulin receptor phosphorylation in the insulinomimetic effects of hydrogen peroxide. *Proc Natl Acad Sci U S A* 84: 8115–8119, 1987.
- Konrad D. Utilization of the insulin signaling network in the metabolic actions of α-lipoic acid—reduction or oxidation? *Antioxid Redox Signal* 7: 1032–1039, 2005.
- 11. Lawrence JC Jr and Larner J. Activation of glycogen synthase in rat adipocytes by insulin and glucose involves increased glucose transport and phosphorylation. *J Biol Chem* 253: 2104–2113, 1978.
- Little SA and de Haen C. Effects of hydrogen peroxide on basal and hormone-stimulated lipolysis in perfused rat fat cells in relation to the mechanism of action of insulin. J Biol Chem 255: 10888–10895, 1980.
- Mahadev K, Wu X, Zilbering A, Zhu L, Lawrence JT, and Goldstein BJ. Hydrogen peroxide generated during cellular insulin stimulation is integral to activation of the distal insulin signaling cascade in 3T3-L1 adipocytes. *J Biol Chem* 276: 48662–48669, 2001.
- May JM and de Haen C. The insulin-like effect of hydrogen peroxide on pathways of lipid synthesis in rat adipocytes. *J Biol Chem* 254: 9017–9021, 1979.
- Mehdi MZ, Pandey NR, Pandey SK, and Srivastava AK. H<sub>2</sub>O<sub>2</sub>-induced phosphorylation of ERK1/2 and PKB requires tyrosine kinase activity of insulin receptor and c-Src. *Antioxid Redox Signal* 7: 1014–1020, 2005.
- Meng TC, Fukada T, and Tonks NK. Reversible oxidation and inactivation of protein tyrosine phosphatases in vivo. *Mol Cell* 9: 387–399, 2002.
- 17. Nomiyama T, Igarashi Y, Taka H, Mineki R, Uchida T, Ogihara T, Choi JB, Uchino H, Tanaka Y, Maegawa H, Kashi-

- wagi A, Murayama K, Kawamori R, and Watada H. Reduction of insulin-stimulated glucose uptake by peroxynitrite is concurrent with tyrosine nitration of insulin receptor substrate-1. *Biochem Biophys Res Commun* 320: 639–647, 2004.
- Perreault M and Marette A. Targeted disruption of inducible nitric oxide synthase protects against obesity-linked insulin resistance in muscle. *Nat Med* 7: 1138–1143, 2001.
- Potashnik R, Bloch-Damti A, Bashan N, and Rudich A. IRS1 degradation and increased serine phosphorylation cannot predict the degree of metabolic insulin resistance induced by oxidative stress. *Diabetologia* 46: 639–648, 2003.
- Rudich A, Tirosh A, Potashnik R, Hemi R, Kanety H, and Bashan N. Prolonged oxidative stress impairs insulin-induced GLUT4 translocation in 3T3-L1 adipocytes. *Diabetes* 47: 1562–1569, 1998.
- 21. Schmid E, El Benna J, Galter D, Klein G, and Dröge W. Redox priming of the insulin receptor beta-chain associated with altered tyrosine kinase activity and insulin responsiveness in the absence of tyrosine autophosphorylation. *FASEB J* 12: 863–870, 1998.
- Shepherd PR, Withers DJ, and Siddle K. Phosphoinositide 3-kinase: the key switch mechanism in insulin signalling. *Biochem J* 333 (Pt 3): 471–490, 1998.

- 23. Sutherland C, Tebbey PW, and Granner DK. Oxidative and chemical stress mimic insulin by selectively inhibiting the expression of phosphoenolpyruvate carboxykinase in hepatoma cells. *Diabetes* 46: 17–22, 1997.
- Werner ED, Lee J, Hansen L, Yuan M, and Shoelson SE. Insulin resistance due to phosphorylation of insulin receptor substrate-1 at serine 302. *J Biol Chem.* 279: 35298–35305, 2004.
- 25. White MF. IRS proteins and the common path to diabetes. *Am J Physiol Endocrinol Metab* 283: E413–E422, 2002.
- 26. White MF. Insulin signaling in health and disease. *Science* 302: 1710–1711, 2003.
- 27. White MF and Kahn CR. The insulin signaling system. *J Biol Chem* 269: 1–4, 1994.

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